

Figure 2: DesignStudio for Custom Probe Design



Manage Targets screen showing Target Region view in DesignStudio. Easily visualize genomic target regions and probes, and assess coverage and score. Design Summary metrics for the entire project are located on the left sidebar, along with project information and user defined labels for convenient data sorting during the design phase.

TruSeq Custom Enrichment Workflow

The custom enrichment workflow begins with pooled, indexed libraries of up to 12 samples that are denatured into single-stranded DNA (Figure 3A) and then hybridized to biotin-labeled custom oligonucleotide capture probes specific to the targeted region (Figure 3B). Streptavidin beads are added to bind to the biotinylated probes (Figure 3C). Biotinylated DNA fragments bound to the streptavidin beads are magnetically pulled down from the solution (Figure 4D). The enriched DNA fragments are then eluted from the beads and re-hybridized for a second enrichment reaction. After amplification of the enriched regions, the targeted library is ready for cluster generation and subsequent sequencing.

Data Analysis

Sequence data generated from custom enrichment samples are analyzed using the TruSeq Enrichment Analysis Script to generate two sets of statistics: post-alignment and post-CASAVA (Consensus Assessment of Sequence and Variation) analysis. Post-alignment analysis counts the number of reads that overlap any targeted region and defines whether a read falls within a target. Post-CASAVA analysis calculates the coverage at each base within a region. Data are visualized using GenomeStudio® Data Analysis Software to examine the on-target and off-target coverage in a sample.

Optimizing Targeted Resequencing

To maximize the efficiency of targeted resequencing studies and ensure that sufficient coverage is obtained for highly sensitive variant calling, three key factors should be taken into account:

1. Sum length of targeted regions, equaling the total amount of targeted genomics sequence (500 kb–25 Mb)
2. Enrichment efficiency (percentage of reads passing filter and mapping to targeted regions)
3. Distribution of coverage depth for targeted regions

Figure 3: TruSeq Custom Enrichment Workflow

A. Denature double-stranded, indexed DNA library

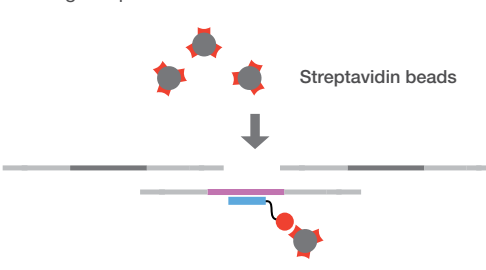
Pooled Sample Library (up to 12-plex)



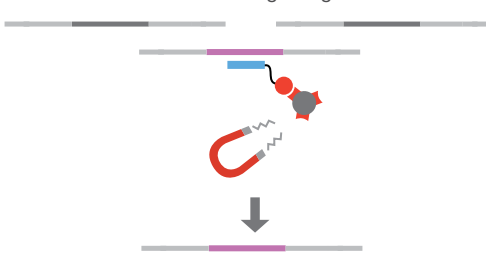
B. Hybridize biotinylated probes to targeted regions



C. Capture of the hybridized biotinylated probes using streptavidin beads

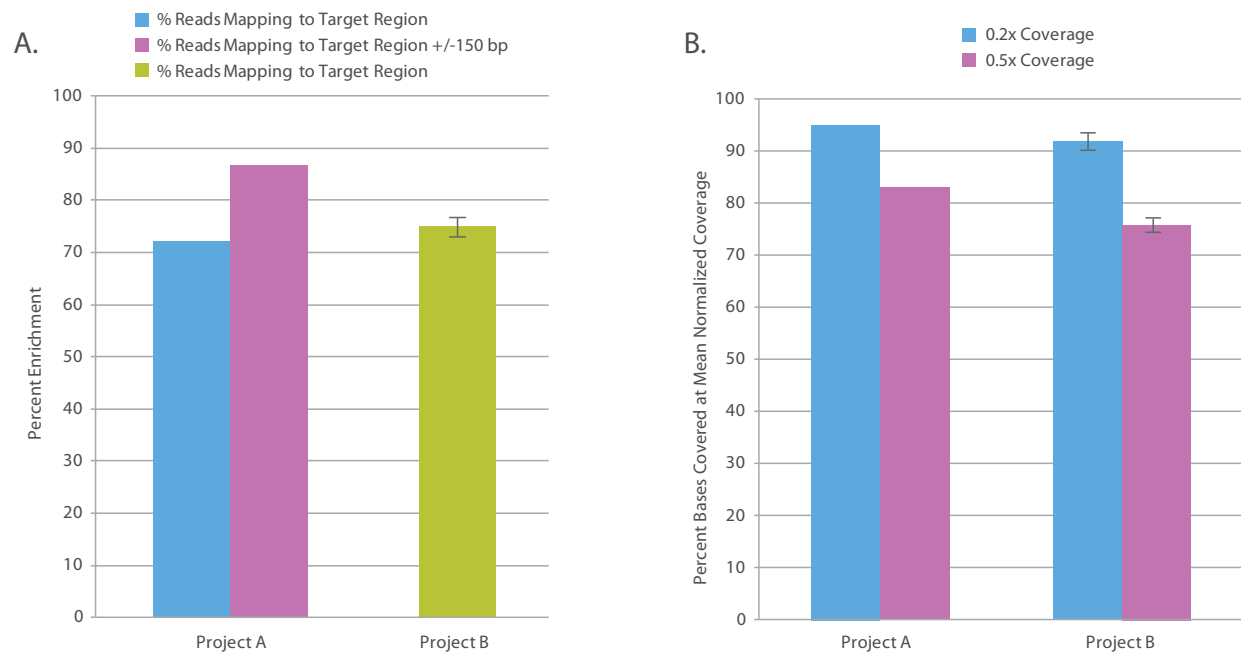


D. Elution of the enriched target regions from beads



The TruSeq Custom Enrichment Kit provides a simple and streamlined in-solution method for isolating and enriching targeted regions of interest. Note that there are two successive rounds of enrichment (B, C, D) in the TruSeq Custom Enrichment workflow.

Figure 4: High Target Specificity and Coverage Uniformity



A. Percent enrichment, defined as reads mapping to target regions out of total reads per run, is shown for two example custom enrichment projects. For single-plex Project A, > 70% enrichment is achieved for reads mapping exactly to target regions (blue). An increase to > 85% is observed when the regions are expanded to +/- 150 bp surrounding the targeted coordinates (purple). For 12-sample multiplexed Project B, the mean percent enrichment averages ~75%, (green) are shown. B. Mean normalized coverage plots for the same two example projects show that > 90% of bases are covered at 0.2x of the mean coverage (blue bars), and > 75% of bases are covered at 0.5x of the mean coverage in both projects (purple bars).

These key parameters and a method for precalculating the amount of sequencing and mean coverage required to fully optimize any targeted sequencing study is described in greater detail in the Optimizing Coverage for Targeted Resequencing Technical Note.²

Data Examples

Two different TruSeq Custom Enrichment experiments were performed following the workflow described in Figure 1. Each project included different target regions, plexities, library sizes, target region sizes, probe interval spacing, and coverage depths (Table 1). Representative enrichment and coverage data are shown in Figure 4. Project A employed single-plex targeting of ~2 Mb total sequence with 20,000 attempted probes. Project B used a 12-plex strategy to target ~1.0 Mb of total sequence with 6,200 attempted probes. Both projects used gel-free TruSeq DNA Library Preparation Kits prior to enrichment, and were sequenced using a Genome Analyzer_{IIx}. In both single- and multiplexed projects, high percent enrichment in targeted and padded regions was achieved, shown in Figure 4. For both projects, mean normalized coverage plots show that > 90% of bases are covered at 0.2x of the mean coverage, and > 75% of the bases are covered at 0.5x of the mean coverage.

Table 1: TruSeq Custom Enrichment Project Details

Detail	Project A	Project B
Unique Bases Targeted	~2.2 Mb	~1.0 Mb
Multiplex Level	1	12
Library Size	350 bp	400 bp
Full Region/Exon	Full Region	Exons
Probe Interval Spacing	Dense	Dense
Reference Sequence	UCSC hg19	UCSC hg19
Total Probes	~20K	~6.2K
Percent Enrichment*	72/87	~75
Percent Bases Covered**	95/83	91/75
Avg. Sequencing Depth†	47x	100x

*Percent enrichment shown as mapped only to target regions/mapped to target regions +/-150 bp (Project A), and to exons (Project B).

**Percent bases covered, shown as mean normalized coverage plots.

†Sequenced on the Genome Analyzer_{IIx}.

